

黄瓜绿斑驳花叶病毒的遗传多样性 与检测技术研究进展

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摘要: 黄瓜绿斑驳花叶病毒(*cucumber green mottle mosaic virus*, CGMMV)是我国重要的植物检疫有害生物, 预防其扩散和传播的关键在于准确、灵敏和快速的检测技术。该文总结分析NCBI数据库中已测定CGMMV分离物的基因组多样性; 比较目前CGMMV主要检测技术的优缺点; 归纳总结当前广泛用于检测CGMMV的逆转录-聚合酶链反应(reverse transcription PCR, RT-PCR)引物, 并针对RT-PCR技术的最新研究进展和存在的一些不足进行讨论; 最后对CGMMV检测技术的未来发展方向进行展望。

关键词: 黄瓜绿斑驳花叶病毒; 检疫; 基因组; 遗传多样性; 检测技术; RT-PCR

Research progresses on genetic diversity and detection technology of cucumber green mottle mosaic virus

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Abstract: Cucumber green mottle mosaic virus (CGMMV) is an important plant quarantine pest in China. The key to prevent its spread lies in accurate, sensitive and rapid detection technology. This paper firstly analyzed and summarized the diversity of the sequenced CGMMV genomes available in the NCBI database; compared the advantages and disadvantages of the current main CGMMV detection technologies; summed up the CGMMV detection primers that widely used in reverse transcription PCR (RT-PCR), and then discussed the latest research progresses and some shortcomings of RT-PCR technology. Finally, this paper looked forward to the future direction of CGMMV detection technology.

Key words: cucumber green mottle mosaic virus; quarantine; genome; genetic diversity; detection technology; RT-PCR

黄瓜绿斑驳花叶病毒(*cucumber green mottle mosaic virus*, CGMMV)自在英国被Ainsworth(1935)首次报道以来, 先后传播到日本(Komuro et al., 1971)、德国(Hentschel, 1975)、伊朗(Rahimian & Izadpanah, 1977)、印度(Raychaudhuri & Varma, 1978)、巴西

(Choudhury & Lin, 1982)、韩国(Lee, 1990)和中国台湾省(Chen & Wang, 1986)等20多个国家和地区。目前, 我国大陆11个省(区、市)74个县(市、区)均发现了CGMMV, 该病毒已被列为全国农业和进境植物检疫性有害生物(沈建国等, 2012; 肖彩利等, 2016),

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主要危害黄瓜、南瓜、西瓜、甜瓜、丝瓜、葫芦、西葫芦和苦瓜等葫芦科作物(Lee et al., 2012; 李立梅等, 2018),引起叶片斑驳、花叶、褪绿、畸形以及果实腐烂、植株矮化和生长缓慢等症状,产量损失严重,防治极其困难,对葫芦科作物的安全生产构成巨大威胁。种子带毒是CGMMV远距离传播的主要途径(宋顺华等, 2015; 2018),病毒颗粒常附着在种子外部表皮上,在花粉、种皮和病苗胚轴中也能发现,为从源头上防止CGMMV随进口种苗传入国内及控制该病毒在我国的进一步扩散蔓延,建立方便、快速、准确和灵敏的CGMMV检测技术是关键环节。本文系统分析NCBI数据库中CGMMV所有分离物基因组的变异情况,比较现有各种CGMMV检测技术的优缺点,重点总结目前应用广泛的反转录-聚合酶链反应(reverse transcription PCR, RT-PCR)扩增引物及该方法应用的最新进展,并对CGMMV检测领域的发展方向进行展望。

1 CGMMV基因组的多样性

通过搜索NCBI数据库,发现CGMMV的基因组序列信息仍在不断增加,显示该病害的扩散仍在继续,越来越多的分离物被获得并进行了基因组测定。通过总结分析NCBI数据库中截止到2021年2月3日的所有CGMMV基因组序列,发现该病毒的基因组大小在6 325~6 440 bp之间,其中CGMMV中国分离物有38株,占该数据库中CGMMV基因组总数的35.19%,其次是欧洲与美洲分离物。这固然可能和我国对该病毒研究上的投入比较大有关,但也从另一个侧面充分显示了该病毒在我国分布广泛,面临着极为严峻的扩散风险。而且分析结果显示该病毒寄主范围广泛,能够侵染我国很多重要的经济作物,包括瓠瓜、黄瓜、西瓜和甜瓜等,防控形势严峻。

本文基于NCBI数据库中所有CGMMV分离物的基因组序列构建了系统发育树,聚类分析发现我国的CGMMV分离物和其他国家的CGMMV分离物整体上可以分为2大类群,第1类群CGMMV分离物的地理来源非常多样,既包括来自亚洲的中国、泰国、日本、韩国、以色列和印度等国家的分离物,也包括来自美洲的美国和加拿大等国家的分离物,还包括来自欧洲的法国、保加利亚、希腊和荷兰等国家的分离物,以及大洋洲的澳大利亚等国家的分离物(图1)。所有来自中国的CGMMV分离物都在第1类群且绝大多数分离物聚合在一起,显示大部分中国分离物可能有相同的地理起源;少数中国CGMMV分离物与其他国家的分离物显示了高度

的基因组同源性,推测可能是随着种子、种苗以及其他植物材料传入我国。第1类群CGMMV分离物的寄主范围也非常广泛,但绝大多数中国分离物都来自于西瓜,少数来自于甜瓜、黄瓜、南瓜和本氏烟等。

第2类群CGMMV分离物的寄主则非常有限,除5个感染黄瓜的分离物外,其他分离物都来自于本氏烟;而且第2类群CGMMV分离物的地理来源也相对有限,除分别有1个分离物来自于美国、科威特和日本外,其他所有分离物都来自于欧洲的荷兰、法国、俄罗斯、德国和西班牙等,其中主要是荷兰(图1)。美国、科威特和日本分离物的寄主都是本氏烟,推测很可能是随着种子、种苗以及其他植物材料由欧洲传入。CGMMV两大类群内部也存在丰富的基因组多样性,例如同样感染西瓜的CGMMV中国分离物聚在不同的亚群,同样感染本氏烟的荷兰分离物也聚在不同的亚群,这可能是由于CGMMV与寄主长期进化引起基因组变异的结果。此外,相对于其他国家,我国的CGMMV分离物寄主范围非常广泛,同一寄主分离物以及不同寄主分离物间都存在着丰富的遗传多样性(图1),这给该病毒的检测增加了极大的挑战,也在一定程度上解释了为何我国有大量RT-PCR检测引物被设计应用。

2 目前主要CGMMV检测技术优劣分析

现有的CGMMV主要检测方法各有优缺点(表1)。其中,生物学检测方法利用疑似病株汁液摩擦接种健康植株或苋色藜、曼陀罗和矮牵牛等鉴别寄主植物,根据发病症状可将CGMMV分为6个株系(Thomas, 1984),该方法简单易行,但存在稳定性较差、工作量大、费时长(宋顺华等, 2014)、同属病毒难区分以及带毒种子长成植株不一定感病等缺点,需结合其他方法才能得到更精确的鉴定结果。电子显微镜通常起辅助鉴定作用,可直观显示病毒粒子的形态和大小、病毒在寄主细胞内的存在状态及受侵染寄主细胞的结构变化等(Debrick, 1973),由于电镜只能观察100 nm以下薄度的样本切片,因此该方法需超薄切片机等精度仪器,对操作人员技术要求严格,难以同时检测大量样本。血清学检测方面应用最多的是酶联免疫吸附测定(enzyme linked immunosorbent assay, ELISA)法(孙玉燕等, 2016),该方法具有操作简单、经济、可批量检测的优点,适合田间大规模鉴定,但需特异性抗体,且由于抗体质量和种子成分易导致假阳性,因此常作为大量样品的初筛手段,阳性样品需经过分子生物学手段鉴定确认。RT-PCR等分子生物学技术的检测灵敏度高于ELI-

SA法和血清学方法,但该方法需要提取RNA,对仪器和操作人员专业技术要求较高,难以用于批量检测。

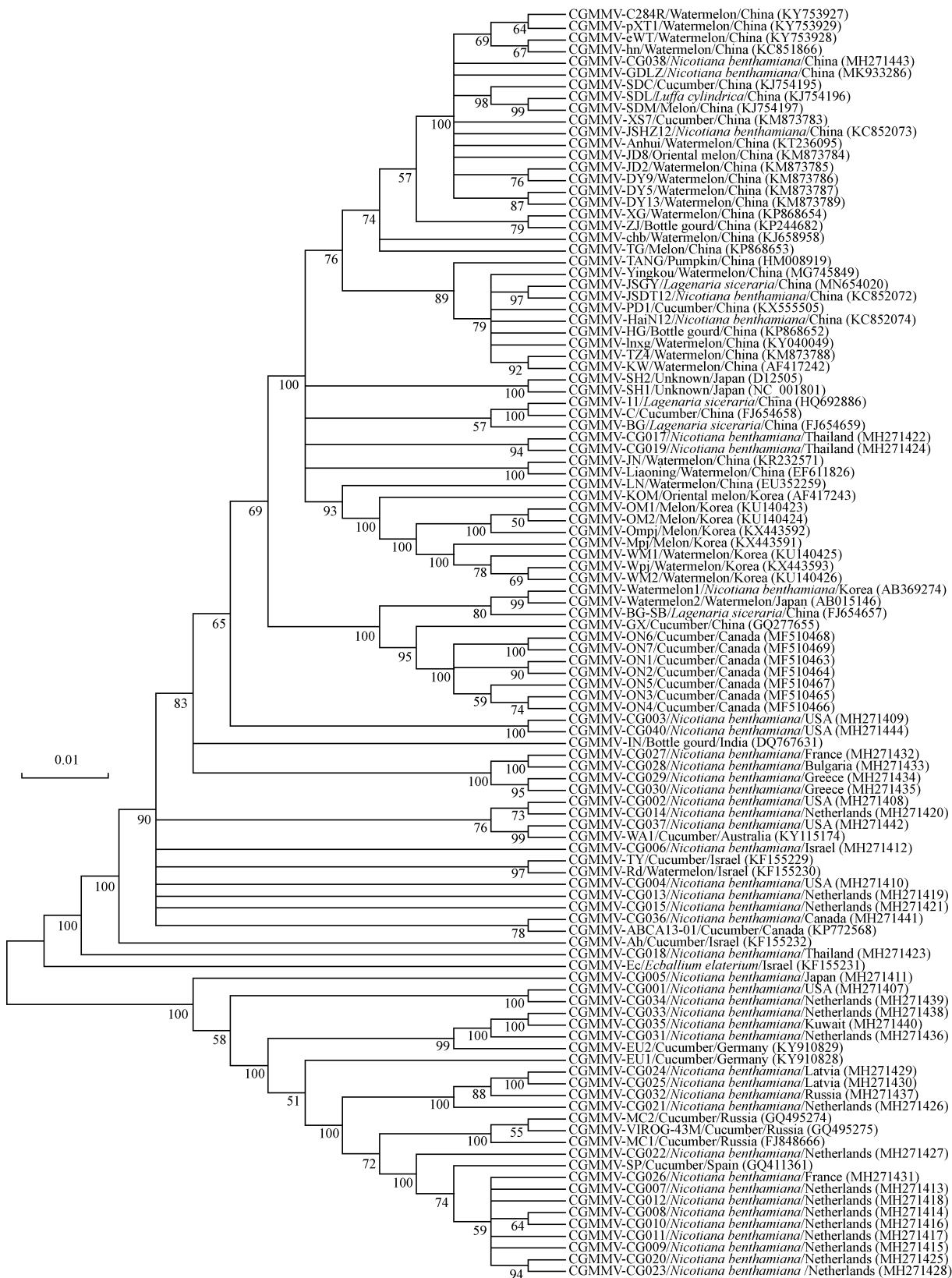


图1 基于CGMMV基因组序列以邻接法构建的系统发育树分析

Fig. 1 Neighbor-joining phylogenetic tree based on CGMMV genome sequences

所有数据均下载自GenBank,每条数据所示为黄瓜绿斑驳花叶病毒-分离物/寄主/来源(GenBank登录号). All data are downloaded from GenBank, each data was presented CGMMV-isolate/host/origin (GenBank accession number).

除上述生物学、电子显微镜、血清学和分子生物学等主要检测技术外,研究人员还建立了其他检测技术,包括荧光原位杂交(孙玉燕等,2016)和核酸斑点杂交(李海明等,2010)等各类杂交检测技术,基因

芯片(张卫东等,2011)和可视化蛋白芯片(胡娟等,2014)等芯片检测技术,以及小RNA深度测序技术(车海彦等,2020)等其他检测方法,但这些技术目前还未普及。

表1 CGMMV检测技术的主要类型及其优缺点

Table 1 Main types of CGMMV detection technology and their advantages and disadvantages

类型 Type	材料/设备要求 Material/equipment requirement	技术要求 Technical requirement	适用场景 Applicable scene	限制因子 Limited factor	检测速度 Detection speed	工作量 Workload	检测成本 Testing cost
生物学 Biology	活体植株 <i>In vivo</i> plants	低 Low	田间或实验室 Field or laboratory	性状表现不稳定 Unstable character	慢 Slow	大 Big	低 Low
血清学 Serology	酶标仪 Microplate reader	中 Medium	田间或实验室 Field or laboratory	抗体质量 Antibody quality	快 Quick	中 Medium	中 Medium
细胞生物学 Cell biology	电子显微镜 Electron microscopy	高 High	实验室 Laboratory	切片质量 Section quality	慢 Slow	大 Big	高 High
反转录PCR RT-PCR	PCR 仪 PCR instrument	高 High	实验室 Laboratory	RNA 质量 RNA quality	快 Quick	中 Medium	中 Medium

3 RT-PCR 检测技术的发展现状及其所用引物

RT-PCR 是目前最为常用的一种方法,具有快

速、准确、特异性强和灵敏度高的优点,国内外相关科研人员主要根据基因组序列上的衣壳蛋白或运动蛋白以及其他保守区序列为靶基因设计了大量的引物,用于CGMMV的检测(表2)。

表2 用于检测CGMMV的RT-PCR引物

Table 2 Primers used for RT-PCR detection of CGMMV

产物 大小 Product size/bp	引物序列(5'-3') Primer sequence (5'-3')	参考文献 Reference
1 053	F: GATGCATCTGTTGCAGAGG/R: TCAACCTCACACACGTAAGAGG	Ali et al., 2004
486	F: ATGGCTTACAATCCGATCAC/R: CTAAGCTTCGAGGTGGTAGC	陈红运等,2006 Chen et al., 2006
-	F: TTTCTGGTGTATGGAACGTA/R: CTCGAAACTAACGTTTCG	任小平,2007 Ren, 2007
715	F: GGCTTACAATCCGATCAC/C/R: CTAAGCTTCGAGGTGGTAGCCT	赵世恒等,2007 Zhao et al., 2007
662	F: ATGGCTTACAATCCGATCAC/R: TGCCCCCTACCCGGGGAAAAG	李红霞等,2007 Li et al., 2007
493	F: ATGGCVFACAATCCGATCAC/R: CTAAGCTVFCGAGGTGGTAGC	白静,2007 Bai, 2007
650	F: CGTGGTAAGCGGCATTCTAACCTC/R: CCGCAAACCAATGAGCAAACCG	黄静等,2007 Huang et al., 2007
486	F: ATGGCTTACAATCCGATCAC/R: CTAAGCTTCGAGGTGGTAGC	秦碧霞等,2008 Qin et al., 2008
280	F: GTTAGTGCTCTTATGTTCCCGTCAGG/ R: TCAGCAGTCGTAGGATTGCTAGGATCTA	邓丛良等,2008 Deng et al., 2008
660	F: ATGGCTTACAATCCGATCAC/R: TGCCCCCTACCCGGGGAAAAG	
500	F: ATGGCTTACAATCCGATCAC/R: CTAAGCTTCGAGGTGGTAGC	
486	F: ATGGCTTACAATCCGATCAC/R: CTAAGCTTCGAGGTGGTAGC	周玲玲等,2008 Zhou et al., 2008
524	F: GAAGAGTCCAGTCTGTTTC/R: ACCCTGAAACTAACGTTTC	张永江等,2008 Zhang et al., 2008
-	F: TTGCATGCTGGCCCCCTACCCGGGGAAAAG/ R: CCGAATTCATGGCTTACAATCCGATCAC	Chen et al., 2008
-	F: GCATAGTGTCTCCGTCAC/R: TGCAGAATTACTGCCATAGAAAC	
284	F: GTTAGTGCTCTTATCTCCCGTCAGG/ R: TCAGCAGTCGTAGGATTGCTAGGATCTA	邓丛良等,2009 Deng et al., 2009
653	F: CGTGGTAAGCGGCATTCTAACCTC/R: CCGCA AACCAATGAGCAAACCG	黄静等,2009 Huang et al., 2009
700	F: CGATCCATGGCTTACAATCCGATCAC/ R: TTGTCGACTGGCCCCCTACCCGGGGAA	黄峰等,2009 Huang et al., 2009
675	F: CGTGGTAAGCGGCATTCTAA/R: AACGTCACACGTGAGAGGTT	李小妮等,2009 Li et al., 2009
1 109	F: CGACGATGCAATCCACGAAT/R: AACACAGGACCCTGAGGAA	

续表2 Continued

产物 大小 Product size/bp	引物序列(5'-3') Primer sequence (5'-3')	参考文献 Reference
1 100	F: GTTTAATTTATAATTAAACAAACAACAAACAA/ R: GCTCGCTTCAGGAAACCAACACAG	陈红运等, 2009 Chen et al., 2009
2 400	F: AAAGCCACGTTCTGTGTGGT/R: GTTCTGCATTAATTGCTATTG	
2 500	F: GCGGATGTGAAAAGGTCATCA/R: GCGGTACCTGTGAAGCAACTAG	
700	F: ATGGCTTACAATCCGATCAC/R: TGGGCCCTACCCGGGGA	
663	F: ATGGCTTACAATCCGATCAC/R: TGGGCCCTACCCGGGAAAG	孙宁等, 2010 Sun et al., 2010
487	F: ATGGCTTACAATCCGATCAC/R: CTAAGCTTOGAGGTGGTAGC	
480	F: CTTACAATCCGATCACACCTAG/R: CTAAGCTTCGAGGTGGTAGC	尚海丽等, 2010 Shang et al., 2010
446	F: GCAAACTTA TTGCCGTTAGTGC/R: GCTTCGAGGTGGTAGCCTCT	阚春月等, 2010a Kan et al., 2010a
158	F: TGCGGGTTTACGCTTTC/R: CAATGACCCTATTACCGCTATCC	
644	F: GCGGCATTCTAACCTCCAATCGG/R: CAAACCCAATGAGCAAACCGTTCG	吴鑫等, 2010 Wu et al., 2010
654	F: CGTGGTAAGCGGCATTCTA/R: CCGCAAACCAATGAGCAAACCG	廖富荣等, 2010 Liao et al., 2010
450	F: CGGGATCCATGGCTTACAATCCGATCACAC/R: GTCGACCTAACGCTTCGAGGTGGTAGCC	李海明等, 2010 Li et al., 2010
591	F: CACCTTATGTCACATTGTTG/R: GTGATCGGATTGTAAGCCATC	
417	F: GTGCTCTTATGTTCCCGTCAG/R: ACCAGACTACCGAAAACGCG	张卫东等, 2011 Zhang et al., 2011
486	F: TCCCCCGGGATGGCTTACAATCCGATC/R: CGGGATCCCTAACGCTTCGAGGTGGTAGC	李海明等, 2011 Li et al., 2011
795	F: AACTGCAGATGCTCTAACGTTAACGCTTCG/R: CGAGCTCCTAGCTGTGATCCGATTGTAAG	吴会杰等, 2011 Wu et al., 2011
482	F: TGGCTTACAATCCGATCACAC/R: AGCTTCGAGGTGGTAGCCT	
1 032	F: YGGYTYATTGTTAACCTG/R: YTGAGTYTGBAARGCGGT	
524	F: GAAGAGTCCAGTTCTGTTTC/R: ACCCTCGAAACTAACGTTTC	郑海刚和陈启建, 2011 Zheng & Chen, 2011
650	F: CGTGGTAAGCGGCATTCTAACCTC/R: CCGCAAACCAATGAGCAAACCG	沈建国等, 2012 Shen et al., 2012
600	F: TAAGCGGCATTCTAACACC/R: TCGATTAAAGTGAAC	刘颖等, 2012 Liu et al., 2012
1 109	F: CGACGATGCAATCCACGAAT/R: AACACAGGACCGTTGAGGAA	商明清等, 2012 Shang et al., 2012
1 475	F: CGACGATGCAATCCACGAAT/R: AACGTCACACGTGAGAGGTT	邵坤彦等, 2012 Shao et al., 2012
309	F: CGTGGTAAGCGGCATTCTAA/R: AACACAGGACCGTTGAGGAA	赵慧茹等, 2013 Zhao et al., 2013
675	F: CGTGGTAAGCGGCATTCTAA/R: AACGTCACACGTGAGAGGTT	蒋庆琳和周国辉, 2014
748	F: ATCAACAACCAACGTGACGC/R: TGCACATCATCACCATCGAC	Jiang & Zhou, 2014
690	F: ATCAACAACCAACGTGACGC/R: CTATGAAGAGCAGGTCCGAT	
638	F: GGTGTATGACGAGGCTGTC/R: TGACATCATCACCATCGAC	
570	F: GGTGTATGACGAGGCTGTC/R: CTATGAAGAGCAGGTCCGAT	
486	F: ATGGCTTACA ATCCGATCACACC/R: CTAAGCTTCGAGGTGGTAGCCTC	王云等, 2014 Wang et al., 2014
976	F: CTAATTATTCTGCTGGCTGCGGATGC/R: CTTGCAGAAATTACTGCCCATA	Tian et al., 2014
523	F: GAAGAGTCCAGTTCTGTTTC/R: ACCCTCGAAAACGTTTC	Nematollahi et al., 2014
1 944	F: GTTTAATTTTATA ATTAAAC/R: CTCCATATCTCAGTTACATCCA	钟敏等, 2015 Zhong et al., 2015
1 563	F: CATGTGAAGATATCGATCTAACTGAAG/R: CCAAACATCGTAAGATCGACTG	
1 617	F: GATCCACTTACAAGTATAATAGCGGATC/R: GAGTTCTGACTGACACCTTAC	
1 463	F: GAGTGATA AGCGCCTTCCGTAG/R: TCGGCCCTACCCGGGG	
654	F: CGTGGTAAGCGGCATTCTAACCTC/R: CCGCAAACCAATGAGCAAACCG	
875	F: TCTGCGGTGGTCG/R: AAAAGGGGGGGATT	杜江等, 2015 Du et al., 2015
660	F: CCACGAGTTCTGCTTAATGCTG/R: TTTGCTAGGCGTGATCGGATTGT	任春梅等, 2015 Ren et al., 2015
850	F: CCACGAGTTCTGCTTAATGCTG/R: TTTGCTAGGCGTGATCGGATTGT	严丹侃等, 2016 Yan et al., 2016
654	F: CGTGGTAAGCGGCATTCTAACCTC/R: CCGCAAACCAATGAGCAAACCG	潘亚南等, 2016 Pan et al., 2016
141	F: TTATTGCGTTAGTGTCTTATG/R: CTACGACAGACGAGGGTAA	Lee et al., 2016
523	F: CCACGAGTTCTGCTTAATGCTG/R: TTTGCTAGGCGTGATCGGATTGT	张艳超等, 2017 Zhang et al., 2017
486	F: CGGAATTCATGGCTTACAATCCGATCACAC/R: CCCTCGAGCTAACGCTTCGAGGTGGTAGCCT	刘锦等, 2017 Liu et al., 2017
243	F: CTTCCCGTCAGGACTTTACTT/R: ATCTACAAACCTCAATGACCTA	Wang et al., 2017
635	F: ATCCCTCGTGCCTGTCAAGT/R: GATCGGATTGTAAGCCATCTC	杨柳等, 2018 Yang et al., 2018

续表2 Continued

产物 大小 Product size/bp	引物序列(5'-3') Primer sequence (5'-3')	参考文献 Reference
874 F: TTGACGGAGTGCCGGTTGTGG/R: AGCGAATTCTGCATTAATTGCTAT		Li et al., 2018
654 F: CGTGGTAAGCGGCATTCTAACCTC/R: CCGCAAACCAATGAGCAAACCG		周莹等, 2019 Zhou et al., 2019
1 004 F: TCCTTGACTTAGAGGTCG/R: TGGGCCCTACCCGGGAA		刘美等, 2019 Liu et al., 2019
- F: ACCCTGAAAACTAAGCTTC/R: ACCCTGAAAACTAAGCTTC		Sui et al., 2019
540 F: ACAGTCATTGATGCTCCG/R: GGTGACAACCATAACCAGGCT		Bi et al., 2019
246 F: CCTCCACCAACCA ATACGCT/R: GTCATTACTGAATTCAACA		
756 F: TGGTGCTACTGTTGCTCTGG/R: GAAGCGAAACGAAGATAGGC		
777 F: GAATCCCTCGTGCCTGTCAAG/R: GGTAAGGCAGACAGGGACTCGC		姜军等, 2020 Jiang et al., 2020
890 F: CCACGAGTTGTTCTAAT/R: TTTGCTAGGCGTATGGATTGT		李正刚等, 2020 Li et al., 2020
771 F: GCGAGGGTCAATTGGAAGA/R: CTGATCGACCTTCCGCAT		车海彦等, 2020 Che et al., 2020

对于CGMMV的检测,普通RT-PCR法的检测灵敏度较高,操作比较简单,但RNA易降解,影响检测结果,现已在普通RT-PCR法基础上研发出不用提取RNA且灵敏度更高的试管捕捉RT-PCR法、免疫捕捉RT-PCR法和免疫磁珠RT-PCR法(表3)。试管捕捉RT-PCR法直接以CGMMV提取液包被PCR管,不需要使用昂贵的特异性抗体,操作更简便,且成本较低,但其检测灵敏度低于免疫捕捉RT-PCR法(沈建国等,2012);免疫捕捉RT-PCR法结合了ELISA法捕捉抗原与RT-PCR法特异性反转录的特点(宋顺华等,2014),通过抗体特异性吸附CGMMV的病毒粒子,降低植物组织PCR抑制物质在CGMMV检测中的抑制作用,特异性强,结果更可

靠,对CGMMV的检测灵敏度是普通RT-PCR法的2倍(阚春月等,2010a,b);免疫磁珠法是将免疫学反应的高度特异性与磁珠特有的磁响应性相结合的一种新免疫学技术,单克隆抗体包被的免疫磁珠法在检测感染CGMMV黄瓜种子时比普通免疫捕捉RT-PCR法的灵敏度高4~8倍(阚春月等,2010a,b),但这2种免疫学方法都需要高灵敏度、高特异性的抗体(表3)。此外,通过同时设计多对不同引物,建立了检测瓜类单粒种子携带CGMMV的双重RT-PCR法和双重免疫捕捉RT-PCR法以及巢式双重免疫捕捉RT-PCR法,最后一种方法要比前面2种方法的灵敏度高10倍(蒋庆琳和周国辉,2014)。

表3 RT-PCR检测技术基础上建立的CGMMV检测技术优缺点

Table 3 The advantages and disadvantages of RT-PCR based technologies for CGMMV detection

类型 Type	灵敏度 Sensitivity	特异性 Specificity	RNA提取 RNA extraction	抗体 Antibody	技术要求 Technical requirement	参考文献 Reference
反转录PCR RT-PCR	高于血清和生物学 >serum and biology	较强 Good	需要 Need	不需要 Don't need	较复杂 Relatively complex	张卫东等,2011 Zhang et al., 2011
免疫捕获RT-PCR IC-RT-PCR	比RT-PCR高2倍 2-fold higher than RT-PCR	强 Strong	不需要 Don't need	需要 Need	简单 Simple	阚春月等,2010a,b Kan et al., 2010a,b
试管捕获RT-PCR TC-RT-PCR	高于RT-PCR,低于免疫捕捉RT-PCR <IC-RT-PCR, >RT-PCR	较强 Good	不需要 Don't need	不需要 Don't need	简单 Simple	沈建国等,2012 Shen et al., 2012
实时荧光定量PCR Real-time qPCR	比RT-PCR高100倍 100-fold higher than RT-PCR	强 Strong	需要 Need	不需要 Don't need	复杂 Complex	邓从良等,2008 Deng et al., 2008
免疫磁珠RT-PCR IP-RT-PCR	比RT-PCR高4~8倍 4~8-fold higher than RT-PCR	强 Strong	不需要 Don't need	需要 Need	简单 Simple	阚春月等,2010a,b Kan et al., 2010a,b
纳米磁珠RT-PCR MNP-RT-PCR	比RT-PCR低10倍 10-fold lower than RT-PCR	较强 Good	不需要 Don't need	不需要 Don't need	复杂 Complex	邓从良等,2008 Deng et al., 2008
细菌磁RT-PCR BMP-RT-PCR	比RT-PCR高10倍 10-fold higher than RT-PCR	较强 Good	需要 Need	不需要 Don't need	复杂 Complex	孙宁等,2010 Sun et al., 2010

RT: Reverse transcription; IC: immunocaptured; TC: tube capture; qPCR: quantitative PCR; IP: immunomagnetic particle; MNP: magnetic nanoparticle; BMP: bacterial magnetic particles.

纳米磁珠 RT-PCR 法利用纳米磁珠的富集效应吸附 CGMMV 病毒粒子直接进行 RT-PCR 检测, 高效快速, 灵敏度比普通 RT-PCR 法低, 但省去了 RNA 抽提过程和病毒抗体的使用, 避免了植物寄主中多糖及一些酚类物质对病毒 RT-PCR 反应的影响, 简化了操作步骤, 缩短了检测时间, 节约了检测成本(邓丛良等, 2008), 但对检测人员的专业技术要求很高, 故未能有效推广使用。细菌磁 RT-PCR 法是利用细菌磁颗粒对植物病毒的非特异性吸附作用提取 CGMMV 的 RNA 进行 RT-PCR 检测(孙宁等, 2010)。实时荧光定量 PCR 法检测 CGMMV 是将 PCR 技术和荧光测定相结合, 与常规 PCR 相比, 具有特异性强、灵敏度高、定量检测、结果准确、产物不需要电泳等优点, 但需要昂贵仪器及试剂, 检测成本较高, 因此不常用作常规检测(表 3)。

4 展望

基因组分析揭示了 CGMMV 基因组具有丰富的多样性, 而目前使用的检测技术各有优缺点, 未来有必要在以下几个方面予以进一步加强。

首先, 开发免疫试纸条等适用于现场的检测技术。由于试纸条不需 PCR 仪等精密仪器, 只要将样品滴在试纸条上, 反应 5~10 min 即可显示结果, 操作简单, 技术要求低, 适用于农户等非专业人员和设备简单的基层检疫部门大规模普筛。目前, 国内针对 CGMMV 先后开发了磁免疫层析试纸条(魏梅生等, 2014)和荧光纳米颗粒试纸条, 但都受制于各种因素没有得到大面积推广。而浙江大学通过单抗制备的胶体金试纸条由于便捷快速、特异性强、检测灵敏度高等优点受到农业农村部的关注, 正在上海市和宁夏回族自治区等地区推广应用。其他适用于现场的技术还有逆转录环介导等温扩增(reverse transcriptase loop-mediated isothermal amplification, RT-LAMP)技术(Li et al., 2013; Hasiów-Jaroszewska et al., 2019; Sui et al., 2019)和逆转录重组酶聚合酶扩增(reverse transcription-recombinase polymerase amplification, RT-RPA)技术(Jiao et al., 2019; Zeng et al., 2019)等。

其次, 努力实现一纸多元检测。同一种植物可能感染多种有害微生物, 例如西瓜和甜瓜种子上经常同时存在瓜类果斑病菌 *Acidovorax avenae* subsp. *citrulli* 和 CGMMV, 这就产生了多元检测的需求, 而目前已开发的试纸条都具有一对一检测功能, 为保证检测的全面性, 使用者往往需要购买多种类的试

纸条, 造成了较大的成本负担, 开发出多元检测试纸条将极大地简化流程, 节省操作人员时间。

最后, 要着手构建种传病毒的分级综合检测体系。应将胶条金试纸条用于基层样品大规模初筛或常规检测, 如有阳性样品需再经过实验室 RT-PCR 技术或其衍生的免疫捕捉 RT-PCR/磁珠 RT-PCR 技术验证确诊, 构建既快速、简便, 又准确、灵敏的多级综合检测体系, 降低种传病毒的漏检率, 为阻止 CGMMV 的传播和蔓延提供技术支撑。

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